REVIEW

Cell cycle and Alzheimer’s disease: studies in non-neuronal cells

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Summary
The most common cause of dementia in the elderly is Alzheimer disease (AD). In Europe, AD is a leading cause of death. The prevalence of this disease in developed countries is increasing because of very significant shifts in life expectancy and demographic parameters. AD is characterized by progressive cognitive impairment, resulting from dysfunction and degeneration of neurons in the limbic and cortical regions of the brain. Two prominent abnormalities in the affected brain regions are extracellular deposits of β-amyloid, and intracellular aggregates of tau protein in neurofibrillary tangles. The role of these features in AD pathogenesis and progression is not yet completely elucidated. Research over the last decade has revealed that the activation of cell cycle machinery in postmitotic neurons is one of the earliest events in neuronal degeneration in AD. Here we summarize evidence to support the hypothesis that cell cycle alterations occur in cells other than neurons in AD sufferers. Immortalized lymphocytes from AD patients have show an enhanced rate of proliferation associated with G1/S regulatory failure induced by alterations in the cyclin/CDK/pRb/E2F pathway. In addition, these cells have a higher resistance to serum deprivation-induced apoptosis. These neoplastic-like features, cell cycle dysfunction and impaired apoptosis can be considered systemic manifestations of AD disease.

Key words: Alzheimer’s disease; lymphocytes; cell cycle; cell survival; p27; p21; calmodulin; PI3K/Akt; ERK1/2

INTRODUCTION

Alzheimer's disease (AD) is a progressive neurodegenerative disorder affecting aged people; AD prevalence is approximately 1% between 65 and 69 years and is higher than 50% in individuals above 95 years. The predominant clinical manifestation is memory loss, but a number of other changes in brain functioning, including impairments in language and visual-spatial skills, and disorientation also characterize this disease. With increasing life expectancy across the world, dementia is a rapidly growing socioeconomic and medical problem. The hallmarks for AD are β-amyloid plaques, neurofibrillary tangles, and regionalized neuronal loss. However our understanding of the role that these features of AD play in the etiology of the disease remains incomplete.

AD is extremely complex and genetically heterogeneous. The majority of patients with the so-called “sporadic” disease exhibit clinical signs during the seventh decade, whereas individuals with inherited AD (FAD) often become demented in mid...
life. To date autosomal dominant FAD has been linked to the presence of mutations in the genes APP, PS1, and PS2, which are genes encoding the amyloid precursor protein (APP), located in chromosome 21 or the presenilins PS1 or PS2 in chromosomes 14 and 1 respectively. The late-onset or sporadic AD has been associated with genetic factors modifying the risk of suffering AD. Notably, the risk of apoE allele type is a susceptibility locus with apoE4 showing a dose-dependent contribution to AD (Strittmatter et al. 1993). Various methods of genetic analysis indicate that additional genes predisposing to AD exist. A complete list of susceptibility genes associated with increased risk for AD can be found in http://alzgene.org (Bertram et al. 2007).

CELL CYCLE AND AD

Despite progress in uncovering many of the factors that contribute to the etiology of this disease, the cause of neuronal death is largely unknown. One promising theory to explain neurodegeneration in AD is that neuronal loss could be associated with cell cycle disturbances. This hypothesis receives support from evidence that a variety of cyclins and cyclin-dependent kinases (CDKs) are up-regulated in the brain of AD patients (Nagy et al. 1998), suggesting that elements that normally control cell cycle progression in proliferating cells may modulate neuronal death as well. Hippocampal and selected cortical neuronal populations in AD exhibit phenotypic changes characteristic of cells re-entering the cell division cycle (Arendt 2003), and it has been further demonstrated that a significant fraction of the hippocampal pyramidal and basal forebrain neurons have fully or partially replicated four independent loci of three different chromosomes (Yang et al. 2001). The successful duplication of DNA indicates that some neurons had completed the S phase of cell cycle (Mosch et al. 2007). These anomalies were not found in unaffected regions of AD brains or in the hippocampus of non-demented age-matched controls. Recent work has shown that cell cycle-related events are present many months before the appearance of plaques or signs of inflammation in brain of murine models of AD (Yang et al. 2006), as well as in brain of patients with Mild Cognitive Impairment (MCI) (Ueberham and Arendt 2005). MCI is a risk stage for development of AD within the next 3–5 years. Therefore, it appears that cell cycle dysfunction is implicated in disease onset and early development and it seems to confer selective vulnerability to neurons (Webber et al. 2005). Importantly, most of the AD pathological features, including β-amyloid, tau, presenilin mutations, oxidative stress, dystrophic neuritis, DNA damage and aneuploidy are somehow related to cell cycle control, providing a link between cell cycle disturbances and neurodegeneration (Zhu et al. 2007, Żekanowski and Wojda 2009). A causal relationship between cell cycle re-entry and neurodegeneration has been recently reported in a transgenic mouse model in which conditional and neuron-specific expression of the proto-oncogene c-Myc leads to cognitive deficit and neurodegeneration (Lee et al. 2009a). It is now assumed that adult neurons, rather than staying permanently postmitotic, must constantly keep their cell cycle in check (Herrup and Yang 2007). If control of neuronal cell cycle fails, the consequence is the entrance of neurons into an altered and vulnerable state, often leading to death (Zhu et al. 2007). Based on these findings, some authors have come to consider AD disease as an abortive neoplastic disorder, that is, a disease of the cell cycle, and thus, the knowledge of the intimate involvement of cell-cycle checkpoints in molecular pathogenesis of AD might be important for diagnostic purposes and particularly in the search for treatment strategies (see Lee et al. 2009b for a review).

MITOTIC SIGNALS

The search for factors responsible for the formation of neurofibrillary tangles and amyloid deposits has yielded several clues to the hypothesis that cell cycle-related phenomena is implicated in the accumulation of AD pathology. CDKs are involved in the phosphorylation of tau (Weishaupt et al. 2003), the main component of tangles as well as a key protein for cytoskeleton organization that occurs during neurite outgrowth and perhaps in aberrant neuronal sprouting (Webber et al. 2005). β-amyloid has been identified as mitogenic in vitro (Milward et al. 1992). APP may induce the activation of cell cycle proteins in neurons (Neve and McPie 2006). Conversely, cell cycle proteins that normally control the progression of cell cycle at the G1/S checkpoint are present in tangle bearing neurons (McShea et al. 2007). The accumulation of potentially mitogenic growth factors (EGF, bFGF) in diffuse amyloid deposits could represent the trigger that initiates the re-entry of neurons into the cell cycle (McShea et al. 1999). DNA damage induced by oxidative stress has been associated with overexpression of the tumor suppressor protein p53 and cell cycle reentry-induced apoptosis in cultured neurons (Kruman et al. 2004).
p53 could activate cell cycle or apoptosis depending on the success of the DNA repairing process (Żekanowski and Wojda 2009). Other factors, including many of the identified risk factors for Alzheimer’s disease, such as elevated plasma homocysteine levels, ageing, menopause, low thyroid levels, low level prolonged oxidative stress or head injury, can either represent mitogenic signalling for neurons or facilitate cell cycle re-entry in vulnerable neuronal populations (reviewed by Arendt 2003).

CELLULAR AND ANIMAL MODELS

Limitations in the use of the postmortem brain for examining molecular mechanisms underscore the need to develop cell or animal models representative of the pathogenesis that characterize AD. Thus, there has been a strong impetus in the last decade to develop a number of different transgenic mouse models of AD that overexpress the human FAD genes in the context of the mouse. This has proven to be a valuable resource in the study of APP processing and in the exploration and design of disease therapies (Morrissette et al. 2009). The AD mice show microglial activation, astrocytosis, and changes in neuronal cytoskeleton proteins including tau. Many of these model organisms have also been shown to have significant memory deficits. Although most of these mice don’t show significant loss of neuronal bodies, cell cycle-related events appear to occur also in the mouse brain. Recently, it has been demonstrated in four different plaque bearing mice, that neurons in the most vulnerable areas have begun a true cell cycle (Yang et al. 2006).

An alternative strategy to study the pathogenesis of AD is the use of non-neuronal cells from patients. Numerous observations indicate that, while the predominant clinical expression arises from brain, AD has systemic expression at the cellular and molecular levels. Considerable precedent exists for studying AD with peripheral tissues, including lymphocytes, fibroblasts, and platelets (Etcheberrigaray and Ibarreta 2001, Casoli et al. 2008). The use of peripheral tissues complement studies of autopsy samples and provide a useful tool to investigate dynamic processes such as signal transduction mechanisms, oxidative metabolism, etc.

CELL CYCLE DISTURBANCES IN NON-NEURONAL CELLS

Work from our laboratory and others have shown cell cycle disturbances in peripheral cells, such as lymphocytes from AD patients, suggesting that dysfunction of the cell cycle is a more general phenomenon affecting cells other than neurons. Immortalized lymphocytes from AD patients showed altered response to mitogenic stimulation relative to control subjects (Urcelay et al. 2001, Nagy et al. 2002). Fibroblasts collected from AD patients also show an aberrant cell cycle-dependent Ca2+ response (Tatebayashi et al. 1995). We, and others have also found failure of the G1/S transition checkpoint, similar to that reported in AD brain, in lymphocytes from AD subjects (Nagy et al. 2002, de las Cuevas et al. 2003), and interestingly in MCI patients as well (Nagy et al. 2002, Zhou and Jia 2010).

We also found that lymphocytes from AD patients were more resistant to serum withdrawal-induced cell death (de las Cuevas et al. 2005), suggesting that control of cell fate depending on the presence or absence of growth stimulatory signals is impaired in peripheral cells from AD sufferers. These features might represent an adaptive response for AD cells that are exposed to chronic stress. It has been considered that susceptible neurons in AD survive for long time in a compromised way by delaying the apoptotic process, a mechanism termed abortosis or abortive apoptosis (Jellinger 2006).

PROLIFERATIVE ACTIVITY OF IMMORTALIZED LYMPHOCYTES FROM CONTROL AND AD PATIENTS

To investigate the distinct cell cycle regulation in AD at the systemic level, we performed a comparative study on cell proliferation, cell cycle profiles, and expression levels of key cell cycle regulatory proteins in lymphoblasts derived from control and late-onset AD subjects. These lymphoblastoid cell lines, obtained by infecting peripheral blood mononuclear cells with the Epstein Barr virus, retained the cellular response of freshly obtained lymphocytes, to serum addition or withdrawal (Bartolomé et al. 2007, Muñoz et al. 2008a). Lymphoblasts from AD patients exhibited a serum dose-dependent enhanced rate of proliferation compared with cells from normal age-matched controls (Urcelay et al. 2001, de las Cuevas et al. 2003). AD lymphoblasts showed cell cycle progress modifications such as a decrease of cells in G1, and an increased number of cells in S phase, together with altered expression and phosphorylation of several proteins involved in regulation of the G1/S transition check point (de las Cuevas et al. 2003, 2005, Muñoz et al. 2005). AD lymphoblasts showed increased
phosphorylation of the retinoblastoma protein (pRb) and other members of the family of pocket proteins compared with cell lines derived from normal age-matched controls (de las Cuevas et al. 2003, Muñoz et al. 2005). pRb is sequentially phosphorylated by two sets of protein kinases, the cyclinD/CDK4 and cyclin E/CDK2 complexes (Mittnacht 1998). The activity of the latter, was found to be enhanced in AD lymphoblasts (Muñoz et al. 2008a). Furthermore, we demonstrated that the increase in CyclinE/CDK2 activity was not due to changes in the expression levels of either cyclin E or CDK2, but rather, to the decreased levels of the CDK inhibitor p27 found in AD lymphoblasts (de las Cuevas et al. 2003, Muñoz et al. 2005).

Once pRb-related proteins are phosphorylated, the transcription factor E2F is released and activated (Weinberg 1995). Accordingly, nuclear extracts from AD lymphoblasts showed reduced E2F-DNA binding activity as determined by EMSA analysis (de las Cuevas et al. 2005). In contrast the activity of NF-κB was found to be decreased in lymphocytes from AD patients, and was not related to the serum-induced enhanced proliferation, but associated instead with decreased vulnerability of AD cells to serum deprivation (de las Cuevas et al. 2005).

Two different reports have shown that freshly obtained lymphocytes from AD patients are less sensitive to G1/S transition blockers, thus suggesting a failure of the G1/S checkpoint function (Nagy et al. 2002, Zhou and Jia 2010). These authors also found these cell cycle alterations in MCI or mild-AD patients.

Further work focused in delineating the molecular mechanisms underlying the p27 down-regulation in AD cells. It was found that this effect was due to increased p27 degradation. A shorter half-life of the p27 protein was detected in AD lymphoblasts as compared with control cells (Muñoz et al. 2008a). p27 proteolysis is a three-step process that requires phosphorylation at Thr187, recognition by the F-box protein SKP2, ubiquitination, and degradation by the 26S proteasome. Increased phosphorylation of p27 protein at Thr187 in AD cells, rather than changes in the 26S proteasome machinery, seems to account for decreased p27 levels. An inverse relationship between phospho-p27 and p27 content was found, while total proteasome activity and accumulation of ubiquitin-tagged proteins did not change significantly (Muñoz et al. 2008a, b).

Interestingly, the enhanced proliferative activity and changes in cell cycle regulatory proteins, can be modulated pharmacologically by treating AD cells with the anti-inflammatory cyclopentenone 15-deoxy-prostaglandin J2 or simvastatin (Muñoz et al. 2005, 2008b, Sala et al. 2008). Therefore these observations provide a plausible explanation for the reported apparent benefits of these drugs preventing or delaying the clinical features of AD (Stewart et al. 1997, Wolozin et al. 2000).

VULNERABILITY OF CONTROL AND AD LYMPHOBLASTS TO SERUM DEPRIVATION

Lymphoblasts from AD subjects were found to be more resistant to serum withdrawal (de las Cuevas et al. 2005, Bartolomé et al. 2007). In control cells, there was a progressive appearance of cell death after 24 h of serum starvation. However, little cell death, as assessed by decreasing levels of MTT reduction was observed in AD cells even after 96 h of serum deprivation (de las Cuevas et al. 2005, Bartolomé et al. 2007). Selective impairment of the mechanisms involved in cell death has been also reported in fibroblasts from AD patients. The protective mechanism of AD fibroblasts against H2O2 was related to an impairment of cell cycle arrest and a diminished induction of apoptosis (Uberti et al. 2002).

The lower sensitivity of AD lymphoblasts to serum withdrawal was associated with changes in the balance of pro- and anti-apoptotic proteins. Moreover it was shown that the survival of AD cells was accompanied by enhanced p21 content as compared with that of control cells (Bartolomé et al. 2009b). A number of recent studies pointed out that in addition to being an inhibitor of cell proliferation, p21 may protect cells from apoptosis (Gartel and Radhakrishnan 2005). For example, it has been reported that up-regulation of p21 blocked the oxidative stress-induced death of human myeloma U266 cells (Kim et al. 2001) and that inducible expression of exogenous p21 render glioblastoma cells resistant to chemotherapy drugs (Ruan et al. 1998). Thus the increase in p21 cellular content in AD lymphoblasts may confer these cells a survival advantage.

SIGNALLING PATHWAYS AND MEDIATORS INVOLVED IN INCREASED PROLIFERATION AND SURVIVAL OF AD LYMPHOCYTES

Since dysregulation of calcium homeostasis is among the major cellular alterations in AD (Thibault et al. 2007) we investigated whether alterations in the
major cellular Ca²⁺-binding protein, calmodulin (CaM) were involved in the altered cellular response of AD lymphoblasts. We found that two structurally unrelated antagonists of CaM, like calmidazolium (CMZ) and W-7, inhibited the proliferation of lymphoblasts exclusively from AD patients (Urcelay et al. 2001, de las Cuevas et al. 2003).

The CaM antagonists were also able to revert the resistance of AD lymphoblasts to cell death induced by serum deprivation (de las Cuevas et al. 2005, Bartolomé et al. 2007). Therefore, CaM seems to play a pivotal role in transmitting proliferative/survival signals from the plasma membrane to the nucleus. Whether CaM contributes to cell proliferation or apoptosis may depend on cellular CaM levels and/or activity, as well as the presence of growth-stimulatory signals.

The combination of CaM antagonist and specific inhibitors of intracellular pathways potentially implicated in the regulation of cell proliferation and apoptosis, revealed the interaction of Ca²⁺/CaM with PI3K/Akt and ERK1/2 pathways in the presence or in the absence of serum, respectively. Table I shows how, in contrast to the selective effect of CMZ, preventing the enhanced stimulation of AD cells, Ly294002, the inhibitor of PI3K/Akt, decreased proliferation of both control and AD lymphoblasts. SB202190, the inhibitor of p38, and PD98059, inhibitor of ERK1/2 had no effect on cell proliferation. However, treatment of control cells with PD98059 prevented cell death induced by serum starvation (Table I). This inhibitor had no effect in AD cells, but blunted the effects of CMZ inducing apoptosis in these cell lines. PI3K/Akt activity, as assessed by increased Akt phosphorylation, was found to be enhanced in AD cells following serum stimulation, compared with the activity observed in control cells (Muñoz et al. 2008b, Bartolomé et al. 2009a, b). In contrast a reduced sustained phosphorylation at Thr187, and protein degradation of PI3K/Akt have similar effects in p27 activity, as well as the presence of growth-stimulatory signals (Pérez-García et al. 2004). As reported for other cell types (Pérez-García et al. 2004), we were able to observe that CaM binds to the p85α subunit of PI3K (Muñoz et al. 2008b). Therefore, CaM could contribute to PI3K overactivation in AD cells through this mechanism, as association of CaM with the SH2 domain in p85 leads to PI3K activation (Pérez-García et al. 2004). The mechanism(s) by which CaM downregulates the ERK1/2 pathway in AD lymphoblasts is not yet known, thought a mechanism of association of CaMKII with Ras-Raf-1/MEK/ERK1/2 seems likely. This issue is currently under investigation in our laboratory.

In summary, our work revealed a functional relationship between Ca²⁺/CaM and PI3K/Akt or ERKs in serum-induced signalling in immortalized lymphocytes, controlling cell fate (proliferation/death or survival) depending on growth factor availability. The proposed scenario is represented schematically in Fig. 1.

Finally, it is worth mentioning that a deregulation of both PI3K/Akt and ERK1/2 signalling pathways has been reported in AD brains. Increased

p27 residues (Thr157, Thr159), which are important in controlling the nucleo-cytoplasmic traffic of p27 (Liang et al. 2002). The exclusion of p27 from the nucleus would then facilitate its degradation by the proteasome (Muñoz et al. 2008a,b) and thus relieve cyclinE/CDK2 kinase activity from p27 inhibition. In fact, it was observed that pRb hyperphosphorylation, E2F, and cyclinE/CDK2 activities, as well as p27 content, were all affected by CaM antagonists (de las Cuevas et al. 2003, 2005, Muñoz et al. 2008a, b). On the other hand, the Ca²⁺/CaM dependent down regulation of ERK1/2 seems to be the key event in protecting AD cells from serum deprivation-induced cell death (Bartolomé et al. 2007). By decreasing the activity of ERK1/2, it induces the overexpression of p21 protein, which in turn blocks the serum withdrawal-induced apoptosis (Bartolomé et al. 2009b). Interestingly, it was found that the CaM binding proteins CaMKII and calcineurin are not involved in the serum-enhanced proliferation of AD lymphoblasts (de las Cuevas et al. 2003, Muñoz et al. 2008a). However CaMKII seems to play an important role in controlling cell survival under serum deprivation, since either CaM antagonists or the CaMKII inhibitor KN-62 sensitize AD cells to death triggered by the absence of growth stimulatory signals (Bartolomé et al. 2007). CaM antagonists had no effect in control cells, suggesting a threshold for CaM activation as the survival signal (de las Cuevas et al. 2005, Bartolomé et al. 2007, Muñoz et al. 2008a). In fact, higher CaM content was found in lymphoblasts from AD patients (Muñoz et al. 2008b).

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Table I. Influence of signalling pretubation on cell proliferation/cell death.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Control</th>
<th>AD</th>
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<tr>
<td></td>
<td>Inhibition of cell proliferation (%)</td>
<td></td>
</tr>
<tr>
<td>+10% FBS</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>20 μM Ly294002</td>
<td>35 ± 3</td>
<td>41 ± 5</td>
</tr>
<tr>
<td>20 μM PD98059</td>
<td>2 ± 4</td>
<td>8 ± 5</td>
</tr>
<tr>
<td>20 μM SB202190</td>
<td>6 ± 4</td>
<td>5 ± 3</td>
</tr>
<tr>
<td>1 μM CMZ</td>
<td>4 ± 2</td>
<td>30 ± 3*</td>
</tr>
<tr>
<td>1 μM CMZ + 20 μM Ly294002</td>
<td>33 ± 4</td>
<td>36 ± 5</td>
</tr>
</tbody>
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|                                | Cell survival (%) |       |
| –10% FBS                       | 69 ± 1            | 92 ± 5‡ |
| 20 μM Ly294002                 | 71 ± 4            | 87 ± 2 |
| 20 μM PD98059                  | 96 ± 2‡           | 92 ± 9 |
| 10 μM SB202190                 | 66 ± 6            | 87 ± 6 |
| 1 μM CMZ                       | 60 ± 3            | 61 ± 6 |
| 1 μM CMZ + 20 μM PD98059       | 93 ± 6‡           | 87 ± 6 |

Lymphoblasts from control and AD individuals were seeded at an initial density of 1 × 10⁶/ml on day 0, in the presence or in the absence of FBS, and treated with the indicated concentrations of drugs for 3 days. Cell survival was expressed as % of cells at day 0. Values shown are the mean 6 SEM for 4–6 observations.

* p<0.05 significantly different from control cells; ‡ p<0.05 significantly from untreated AD cells.

phospho-Akt (Ser473) has been detected in AD temporal cortex, accompanied by increased levels of phosphorylation of Akt substrates such as GSK3, mTOR, tau and lower levels of p27 (Griffin et al. 2005). Increased phosphorylated p27 (Thr 187) has been also found in AD brains. Importantly, Thr187-p27 shows a considerable overlap with tau-positive neurofibrillary pathology, including neurofibrillary tangles and dystrophic neuritis (Ogawa et al. 2003). On the other hand, ERK1/2 activation has been shown in degenerative neurons in close association with neurofibrillary tangles, suggesting the implication of this pathway in AD pathogenesis (Knowles et al. 1999).

Therefore, peripheral cells from patients may be a potential useful surrogate for diagnosis, prognosis and therapeutic monitoring of AD.

While most of the studies on AD lymphocytes had been focused on detecting disease-specific changes that may serve as biomarkers, the clinical consequences, if any, of the enhanced proliferative response of lymphocytes in AD patients, remain to be established, as well as the role that they may play in the chronic inflammation associated with this disease.

CONCLUSIONS

Cell cycle disturbances are evident in non-neuronal cells from AD patients. While the precise origins of cell cycle alterations are not fully understood, a complex interaction of Ca²⁺/CaM with the PI3K/Akt
Fig. 1. Depiction of the proposed signalling pathways altered in AD lymphoblasts.
In AD cells, increased levels of CaM synergise with serum stimulation and promote overactivation of PI3K/Akt leading to enhanced p27 degradation, and activation of cyclin/CDK/pRb/E2F, therefore favouring, the progression of cells through the cell cycle. In the absence of serum, the Ca^2+/CaM-binding protein, CaMKII, decreases the serum-deprivation induced NF-κB and ERK1/2 activation in comparison with control cells, and increases the cellular content of p21, which then seems to protect AD lymphoblasts from apoptosis induced by the absence of trophic support.

and ERK1/2 pathways seems to be the master regulator of cell survival, controlling cell proliferation or preventing apoptosis depending on growth conditions. Two cell cycle regulatory proteins, the CDK inhibitors, p27 and p21, are ultimately responsible for the enhanced proliferation and increased resistance to cell death, respectively. Whereas downregulation of p27 in a Ca^2+/CaM-dependent manner induces the enhanced proliferative response of immortalized lymphocytes from AD patients, upregulation of p21 seems to help AD cells to escape from serum deprivation-induced apoptosis. The distinct cell cycle and apoptosis control in lymphoblastoid cells from AD patients offer a noninvasive tool for investigating the pathogenesis of AD and suggest a number of molecular targets for potential AD therapies.

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REFERENCES

Arendt T: Synaptic plasticity and cell cycle activation in neurons are alternative effector pathways: the ‘Dr. Jekyll and Mr. Hyde concept’ of Alzheimer’s disease or the yin and yang of neuroplasticity, Prog Neurobiol 71:83–248, 2003.


McShea A, Lee HG, Petersen RB, Casadesus G, Vincent I, Linford N, Funk JO, Shapira RA, Smith MA: Neuronal cell cycle re-entry mediates...