Association of FGF19, FGF21 and FGF23 with carbohydrate metabolism parameters and insulin resistance in patients with chronic kidney disease

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Abstract
Insulin resistance (IR) is characterised by increased gluconeogenesis in the liver and the resistance of peripheral receptors to insulin. Several factors, including IR, type 2 diabetes, new-onset diabetes after transplant (NODAT) and secondary parathyroidism, are related to chronic kidney disease (CKD). These factors are associated with higher mortality due to the increased risk of cardiovascular complications. Many factors have been identified as potential markers of IR in CKD. These factors include fibroblast growth factors (FGFs), a subfamily of endocrine polypeptides. In this study, we examined the association of FGF19, FGF21 and FGF23 with selected parameters related to carbohydrate metabolism and insulin resistance in non diabetic patients with predialysis CKD and in non diabetic patients after renal transplantation.

The study included 108 non diabetic subjects: 40 patients with predialysis CKD, 45 patients with CKD who had undergone renal transplantation, and 23 healthy subjects (control group).

In patients who had undergone renal transplantation, concentrations of FGF23 were increased compared to the control group and patients with predialysis CKD. The highest and lowest FGF19 concentrations were observed in CKD patients and in patients who had undergone kidney transplantation, respectively. This difference was statistically significant. Leptin concentrations were higher in CKD patients compared to the control group and patients who had undergone kidney transplantation. There were no statistically significant differences in adiponectin concentrations, lean body mass or fat tissue mass between the studied groups. HOMA-IR and insulin levels were significantly increased in CKD patients and in patients who had undergone renal transplantation in comparison to the control group.

The results of the study suggest the involvement of FGF in carbohydrate metabolism and insulin resistance in patients with predialysis CKD, as well as a correlation with kidney function.

Keywords: Body composition; Chronic kidney disease; FGF19; FGF21; FGF23; Insulin

Highlights:
• Insulin resistance is characterised by increased gluconeogenesis.
• Chronic kidney disease (CKD) is associated with insulin resistance.
• Fibroblast growth factors are involved in carbohydrate metabolism and insulin resistance in patients with predialysis CKD.

Abbreviations: FGF, fibroblast growth factor; CKD, chronic kidney disease; HDL, high-density lipoprotein; LDL, low-density lipoprotein

Introduction
Insulin is a hormone produced by pancreatic beta cells that plays an important role in carbohydrate, lipid and protein metabolism. Insulin resistance (IR) is characterised by the impaired action of insulin on tissues and organs, and is associated with several diseases. IR may be associated with increased gluconeogenesis in the liver and resistance of peripheral receptors to insulin. As a result, the pancreas produces more insulin, leading to hyperinsulinemia. This phenomenon also occurs in chronic kidney disease (CKD), arising in the early stages of the disease (Fliser et al., 1998; Spoto et al., 2016). IR has a significant impact on the progression of kidney disease, especially...
cardiovascular complications (CV) (Kuršat et al., 2010). IR can persist even after successful kidney transplantation, and can progress to new-onset diabetes after transplant (NODAT) with the initiation of immunosuppressive therapy. Because IR is a modifiable risk factor, its reduction may lower cardiovascular mortality in patients with CKD. The mechanism of IR in CKD is mainly related to peripheral resistance, and is associated with endothelial dysfunction (Voytovich et al., 2006). Many newly discovered markers are potentially related to IR in CKD. Recent research highlighted the use of cytokines and adipocytokines that are known to influence insulin sensitivity as potential biomarkers of IR. These factors include the subfamily of endocrine polypeptides known as fibroblast growth factors (FGF) (Garland et al., 2014; Hui et al., 2018; Kliwer and Mangelsdorf, 2015; Sit et al., 2018; Strowski, 2017; Tanajak et al., 2018). These proteins ameliorate IR through various mechanisms that act to stimulate glucose uptake, and are considered therapeutic targets in diseases characterised by IR including obesity, CKD and diabetes (Fernandes-Freitas and Owen, 2015).

In 1999, FGF19 was first discovered in the human brain, and was subsequently detected in many other tissues. The basic roles of FGF19 are related to bile acid secretion, stimulation of glycogen synthesis and inhibition of gluconeogenesis by inhibiting the expression of the transcription factor peroxisome proliferator-activated receptor-gamma coactivator alpha (PGCo) and its target genes (Fukumoto, 2008; Zhang et al., 2015). Previous studies have reported elevated levels of FGF19 in CKD; however, its role in CKD is not fully understood. In addition, Reiche et al. (2010) observed that the serum FGF19 concentration was 1.5 times higher in haemodialysis patients compared to healthy subjects.

FGF21 is primarily found in the liver and adipose tissue, as well as in the skeletal muscle, heart, kidneys and testes (Fukumoto, 2008; Zhang et al., 2015). FGF21 plays important roles in ketogenesis, gluconeogenesis, weight loss, thermogenesis and transformation of white to brown adipose tissue (Nishimura et al., 2000). FGF21 increases glucose tolerance and insulin sensitivity, and also reduces blood glucose levels. Importantly, an overdose of FGF21 does not lead to hypoglycaemia as with insulin. Recent studies suggest that the FGF21 agonist is a promising therapeutic agent for the treatment type 2 diabetes and obesity (Hu et al., 2018). Furthermore, FGF21 was found to be closely associated with renal dysfunction in subjects with end-stage renal disease (ESRD) (Han et al., 2010). There is also an association between serum FGF21 levels and the rate of cardiovascular events and mortality in hemodialysis patients (Suassuna et al., 2019).

FGF23 is a phosphatonin that is produced in response to phosphate retention in CKD, and therefore plays a role in regulating phosphate serum concentrations. Previous studies have confirmed the association of FGF23 with the progression of kidney disease, inflammation, vascular calcification and left ventricular hypertrophy (Isakova et al., 2018; Liu and Quares, 2007). The involvement of FGF23 in carbohydrate metabolism and IR in patients with CKD is not known.

Both IR and type 2 diabetes are important factors in the progression of CKD to ESRD, and are associated with increased mortality due to cardiovascular complications.

In our previous studies we have indicated that increased concentrations of FGF19 and FGF21 in patients with CKD may be associated with the metabolism of lipids and carbohydrates (Marchelek-Myśliwiec et al., 2019a). Additionally haemodialysis and transplantation resulted in the reduction of FGF19 and FGF21 concentrations in patients with CKD (Marchelek-Myśliwiec et al., 2019b). Therefore, in this study we examined the association of FGF19, FGF21 and FGF23 with selected parameters of carbohydrate metabolism and insulin resistance in non diabetic patients with predialysis CKD and in non diabetic patients after renal transplantation.

Materials and methods

Study groups

The study included 108 non diabetic subjects: 40 patients with predialysis CKD, 45 patients with CKD who had undergone renal transplantation, and 23 healthy subjects as the control group. The inclusion criteria for the control group (C) were as follows: aged over 18 years, no chronic inflammatory diseases (e.g., systemic lupus erythematosus, rheumatoid arthritis), no diabetes, and a glomerular filtration rate (GFR) above 60 ml/min/1.73 m². The following inclusion criteria were applied to the predialysis CKD group: aged over 18 years, no diabetes, no need for haemodialysis or renal replacement therapy, and stable renal function for 3 months prior to enrolment. The exclusion criteria for this group were cancer and/or treatment with steroids at a dose of more than 5 mg/day.

The causes of kidney insufficiency were as follows: hypertension, autosomal dominant polycystic kidney disease, g lumeronephritis, congenital urinary tract malformations and nephrolithiasis. The inclusion criteria for transplant patients (Tx) were the following: renal transplantation at least 2 years before enrolment, no diabetes, constant doses of immunosuppressive drugs for the last 3 months, and standard immunosuppression with calcineurin inhibitors, mycophenolate mofetil and glucocorticosteroids. The exclusion criteria were neoplastic disease and/or bariatric surgery. This study was approved by the Bioethical Commission at the Pomeranian Medical University. All participants provided written informed consent.

Materials

Blood samples for the C, CKD and Tx groups were collected from the peripheral vein (using Sarstedt S-Monovette tubes with clotting activator) after an overnight fast. Whole blood was centrifuged, and the serum samples were frozen at −80 °C until use.

Determination of biochemical parameters

Lipid profile (total cholesterol, HDL and LDL cholesterol, triglycerides), glucose, creatinine, uric acid, urea, albumin, total protein, sodium, potassium, magnesium concentrations were determined in serum using the Architect c8000 analyser (ABBOTT, USA). The GFR was estimated using the CKD-EPI equation (eGFR). The insulin concentration was determined by an immunoenzymatic method (ELISA; uIU/ml; R&D). HOMA-IR was calculated according to the formula (glucose (mg/dl) × insulin (uIU/ml)) / 405 (mg/dl × µIU/ml). The serum concentrations of leptin (ng/ml) and adiponectin (µg/ml) in the samples were determined by ELISA (Bio Vendor, Czech Republic), and the concentrations of FGF19, FGF21 and FGF23 (pg/ml) were determined by ELISA (Cusabio Biotech, Wuhan, China).

Determination of body composition

Measurements of adipose tissue content (BF) and lean body mass (LM) were made using the dual X-ray absorptiometer (DXA) method (Lunar Prodigy Advance, GE Healthcare), and the results are expressed in kilograms.
Statistical analysis
Non-parametric tests were used for statistical analysis of quantitative variables (Shapiro–Wilk test). The Mann–Whitney U test was used for comparisons between two groups, and Spearman’s rank correlation coefficient for the assessment of associations between variables within groups. Values of \( p < 0.05 \) were considered to indicate statistical significance for all analyses.

Results

General characteristics and laboratory data
The baseline characteristics of the study groups are shown in Table 1.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>C ( n = 23 )</th>
<th>Predialysis CKD ( n = 40 )</th>
<th>Tx ( n = 45 )</th>
<th>C vs CKD</th>
<th>C vs Tx</th>
<th>CKD vs Tx</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>Mean ± SD</td>
<td></td>
<td></td>
<td>( p^A )</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Body weight (kg)</td>
<td>53.8 ± 13.2</td>
<td>59.9 ± 8.9</td>
<td>54.2 ± 12.4</td>
<td>0.073</td>
<td>0.99</td>
<td>0.030</td>
</tr>
<tr>
<td>BMI (kg/m(^2))</td>
<td>76.4 ± 15.1</td>
<td>80.7 ± 15.3</td>
<td>75.5 ± 14.3</td>
<td>0.25</td>
<td>0.84</td>
<td>0.11</td>
</tr>
<tr>
<td>GFR (ml/min/1.73 m(^2))</td>
<td>26.6 ± 4.1</td>
<td>28.8 ± 5.0</td>
<td>26.4 ± 3.6</td>
<td>0.29</td>
<td>0.62</td>
<td>0.052</td>
</tr>
<tr>
<td>Creatinine (mg/dl)</td>
<td>91.1 ± 17.1</td>
<td>36.2 ± 13.7</td>
<td>57.9 ± 22.3</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
<td>0.000043</td>
</tr>
<tr>
<td>Uric acid (mg/dl)</td>
<td>0.8 ± 0.1</td>
<td>2.0 ± 0.8</td>
<td>1.5 ± 0.6</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
<td>0.0012</td>
</tr>
<tr>
<td>Urea (mg/dl)</td>
<td>4.5 ± 1.3</td>
<td>5.3 ± 1.1</td>
<td>5.5 ± 1.7</td>
<td>0.027</td>
<td>0.016</td>
<td>0.92</td>
</tr>
<tr>
<td>Albumin (g/dl)</td>
<td>33.3 ± 7.8</td>
<td>73.0 ± 27.1</td>
<td>53.7 ± 23.6</td>
<td>&lt;0.0001</td>
<td>0.00078</td>
<td>0.00061</td>
</tr>
<tr>
<td>Total protein (g/dl)</td>
<td>3.6 ± 0.7</td>
<td>4.0 ± 0.9</td>
<td>3.2 ± 0.4</td>
<td>0.16</td>
<td>0.016</td>
<td>0.00035</td>
</tr>
<tr>
<td>Sodium (mmol/l)</td>
<td>6.4 ± 1.3</td>
<td>7.2 ± 1.5</td>
<td>6.0 ± 0.9</td>
<td>0.018</td>
<td>0.25</td>
<td>0.00019</td>
</tr>
<tr>
<td>Magnesium (mmol/l)</td>
<td>4.4 ± 0.3</td>
<td>4.5 ± 0.5</td>
<td>4.4 ± 0.5</td>
<td>0.12</td>
<td>0.70</td>
<td>0.049</td>
</tr>
<tr>
<td>Glucose (mg/dl)</td>
<td>0.91 ± 0.1</td>
<td>0.87 ± 0.13</td>
<td>0.86 ± 0.10</td>
<td>0.054</td>
<td>0.12</td>
<td>0.85</td>
</tr>
<tr>
<td>CH (mg/dl)</td>
<td>97.2 ± 21.2</td>
<td>100.4 ± 22.4</td>
<td>96.2 ± 12.7</td>
<td>0.16</td>
<td>0.26</td>
<td>0.57</td>
</tr>
<tr>
<td>LDL (mg/dl)</td>
<td>182.7 ± 23.7</td>
<td>188.7 ± 36.0</td>
<td>193.6 ± 36.3</td>
<td>0.76</td>
<td>0.22</td>
<td>0.34</td>
</tr>
<tr>
<td>HDL (mg/dl)</td>
<td>108.9 ± 23.5</td>
<td>117.9 ± 36.0</td>
<td>115.9 ± 31.5</td>
<td>0.43</td>
<td>0.55</td>
<td>0.99</td>
</tr>
<tr>
<td>TG (mg/dl)</td>
<td>39.2 ± 8.2</td>
<td>36.0 ± 8.5</td>
<td>42.1 ± 14.5</td>
<td>0.17</td>
<td>0.90</td>
<td>0.20</td>
</tr>
</tbody>
</table>

\( ^A \) Mann–Whitney U test

C, control group; CKD, non diabetic patients with predialysis chronic kidney disease; Tx, non diabetic patients after renal transplantation; BMI, body mass index; GFR, glomerular filtration rate; CH, total cholesterol; LDL, LDL cholesterol; HDL, HDL cholesterol; TG, triacylglycerols.

Statistically significant results \( (p < 0.05) \) are marked in bold.

The highest FGF19 concentrations were observed in predialysis CKD patients and the lowest in patients who had undergone kidney transplantation (Fig. 1A). The differences were statistically significant. The highest FGF21 concentrations were in predialysis CKD patients; however, the differences were not statistically significant (Fig. 1B). In patients who had undergone renal transplantation, we observed increased concentrations of FGF23 compared to the control group and patients with predialysis CKD (Fig. 1C).

The insulin levels and HOMA-IR values were significantly increased in predialysis CKD patients and in patients who had undergone renal transplantation in comparison to the control group (Fig. 2A, B). Leptin concentrations were increased in predialysis CKD patients compared to the control group and patients after kidney transplantation (Fig. 2C). There were no statistically significant differences in adiponectin concentrations (Fig. 2D), lean body mass or fat tissue between the studied groups (Fig. 3A, B).

Correlations
In predialysis CKD patients and patients who had undergone renal transplantation, FGF19 was negatively correlated with GFR \( (R_S = -0.74 \ p < 0.0001 \text{ and } R_S = -0.65 \ p < 0.0001) \). Moreover, in predialysis CKD patients, FGF21 was negatively correlated with leptin levels (Fig. 4A), positively correlated with BMI, GFR, leptin, adiponectin, BF, LM, glucose, insulin, HOMA-IR) in studied groups.
Fig. 1. Concentrations of fibroblast growth factors 19, 21 and 23 in patients with chronic kidney disease, patients after renal transplantation and controls

Concentrations of FGF19, FGF21, FGF23 are shown on logarithmic scale.

* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs control group
^ $p < 0.05$, ^^ $p < 0.01$, ^^^ $p < 0.001$ vs CKD group
C, control group; CKD, non diabetic patients with predialysis chronic kidney disease; Tx, non diabetic patients after renal transplantation.
Fig. 2. Selected parameters of carbohydrate metabolism in patients with chronic kidney disease, patients after renal transplantation and controls

Fig. 3. Content of body fat and lean mass in patients with chronic kidney disease, patients after renal transplantation and controls
**Fig. 4.** Correlations between fibroblast growth factor 21 and selected parameters of carbohydrate metabolism in patients with chronic kidney disease

Concentrations of FGF21 are shown on logarithmic scale. Spearman rank correlation coefficient is presented with corresponding \( p \)-value.

**Discussion**

In this study, we examined the association between FGF and selected parameters of carbohydrate metabolism and insulin resistance (IR) in patients with predialysis CKD. IR commonly accompanies CKD and worsens its prognosis. In recent years, several new factors have been found to be significantly associated with IR (Degirolamo et al., 2016; Kharitonenkov and Shanafelt, 2009; Liu and Quarles, 2007). In our study, we analysed the FGF subfamily and its relationship to carbohydrate metabolism, IR and body mass composition in a group of patients with predialysis CKD. The FGF19 concentrations were negatively correlated with GFR in the CKD and Tx groups. This confirms the association of FGF19 with renal function.

We did not observe an association between FGF19 concentrations and parameters of carbohydrate or lipid metabolism or HOMA-IR. Animal studies have shown that exogenous administration of FGF19 can reduce body weight and alter fat tissue distribution (Tomlinson et al., 2002). In a study by Hu et al. (2018), FGF19 concentrations were found to depend on the amount of adipose tissue accumulated, and were independent of carbohydrate disorders. In our study, we did not find any correlations between FGF19 and the amounts of adipose tissue or lean body mass.

Previous studies have reported increased FGF21 levels in patients with diabetes, obesity, arteriosclerosis and CKD (Mraz et al., 2009). In our study, FGF21 was negatively correlated with GFR in the Tx group, but this correlation was absent in the predialysis CKD and control groups. Interestingly, the concentrations of FGF21 observed in the CKD group were three times higher than in the control and Tx groups, but this difference did not reach statistical significance.

Attempts to explain the elevated concentrations of FGF21 in CKD are based on three concepts. The first one is a compensatory FGF21 response to metabolic stress, while the second concerns the resistance of target tissues to FGF21. The third hypothesis considers the impaired excretion of FGF21 by the kidneys at low GFR (Anuwatmatee et al., 2019; Stein et al., 2009). In contrast to FGF19, we observed a positive correlation of FGF21 with the HOMA-IR index and insulin concentration in all groups.
FGF21 is a late-acting hypoglycaemic hormone. In an animal model, it was demonstrated that the main mechanism of action of FGF21 in terms of glycaemic control is to increase the sensitivity of target tissues to insulin (Coskun et al., 2008). In humans, a study using an FGF21 analogue showed beneficial effects on the lipid profile, adiponectin concentrations and weight loss (Gaich et al., 2013). Our results suggest that FGF21, regardless of renal function, may be a marker of IR. Lin et al. (2010) showed a positive correlation between FGF21 and the amount of subcutaneous fat in obese people with normal insulin sensitivity. Other studies have confirmed an association of FGF21 with adipose tissue and the production of adiponectin (Zhang et al., 2015). Treatment with FGF21 enhanced both expression and secretion of adiponectin in adipocytes, thereby increasing serum levels of adiponectin (Lin et al., 2013). In our study, FGF21 was positively correlated with the adiponectin concentration in the Tx group and negatively correlated with the leptin concentration in the predialysis CKD and Tx groups. In a study by Asrhi et al. (2016) conducted in an animal model and in vitro on human-derived hepatocarcinoma cells, elevated FGF21 concentrations were observed after leptin administration.

In our study, FGF23 was not correlated with parameters of carbohydrate metabolism and body composition. In a previous study by Fayed et al. (2018), FGF23 was found to be an important predictor of IR in CKD patients. In contrast, Hanks et al. (2015) found that FGF23 was not correlated with IR in a group of patients with CKD. Zaheer et al. (2017) showed that BMI and adipose tissue mass were positively correlated with FGF23 in patients with CKD. In another study performed in subjects with simple obesity, a negative correlation was found between FGF23 and HOMA-IR and insulin concentration (Wojcik et al., 2012).

FGF19, FGF21 and FGF23 are involved in the transmission of a number of signals that maintain homeostasis of the body. FGF concentrations are known to be altered in many common and chronic diseases such as obesity, diabetes and CKD, therefore, FGFs can be considered biomarkers of these diseases. Currently, research is being carried out on the use of FGFs as a therapy in liver diseases, type 2 diabetes and obesity, as well as to treat complications of kidney disease. Further studies are needed in CKD patients to explain the role of the FGF19 subfamily in kidney disease.
Conclusions
The results of this study suggest the involvement of FGFs in carbohydrate metabolism and insulin resistance in patients with predialysis CKD, as well as a correlation with kidney function. FGF19 was found to be negatively correlated with GFR. FGF21 concentrations were positively correlated with insulin levels, HOMA-IR and lean body mass.

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Conflict of interests
The authors declare that they have no conflicts of interests.

References


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